

Paradoxical effects of chemicals in the diet on health

Anthony Trewavas* and Derek Stewart†

In 1992, Block *et al.* [1] published a summary of 200 epidemiological investigations which indicated that a diet that was high in fruit and vegetables cut cancer risks approximately in half. These investigations used conventionally farmed produce that contained traces of synthetic pesticides and mycotoxins as well as an estimated 10 000 secondary products (i.e. natural pesticides). Dietary consumption of fruits and vegetables also reduces risks of cardiovascular disease, cataracts and brain dysfunction. Before genetic manipulation is undertaken to elevate or diminish any individual constituent of fruits and vegetables, the contribution of each of these constituents to health must be better understood, as in many cases their effects on health can be paradoxical.

Addresses

*Institute of Cell and Molecular Biology, Mayfield Road, University of Edinburgh, Edinburgh EH9 3JH, UK

†Quality Health and Nutrition Programme, Genes to Products Theme, Scottish Crop Research Institute, Invergowrie, Dundee DD2 5DA, UK

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Abbreviations

EPA Environmental Protection Agency
MTD maximum tolerated dose

Introduction

Ever since the discovery of vitamins, there has been intense interest in the effects of food constituents on human health. The requirements for health of a protein balance containing all the essential amino acids, some unsaturated lipids, minerals and carbohydrates were established early on. But food also contains large numbers of other small molecules, plant secondary products, agricultural and food-processing synthetic chemicals and fungal by-products. With one third of premature human cancers seemingly related to diet [2], intense investigation is now focussing on the health effects of some of the small molecules in our diets. This short review focuses on the increasingly complex situation that is emerging in which even the concept of toxins, long established in the toxicology literature, is likely to undergo a paradigm shift. Many compounds have both health-improving and health-debilitating effects depending solely on the dose.

Natural carcinogens (pesticides)

Fruits and vegetables synthesise a great variety of secondary products that have evolved to control pest damage. Environmental conditions and/or predation can increase the synthesis of these secondary products. Of the 127 plant secondary products that have been tested in rodents (out of an estimated total of 10 000 in plants), more than half are carcinogenic at the maximum tolerated dose (MTD) [2]. However, some of the compounds that have been identified as carcinogenic have very different effects at lower concentrations. For example, quercetin is now recommended as a potent antioxidant that has anticancer properties [3,4]. Limonene and perillyl alcohol are now recognised anti-cancer agents [5]. Caffeic acid at normal levels of human consumption reduces cancer rates to below those in controls [6]. Allyl isothiocyanate is a potent inhibitor of tumour development [7]. And, in moderation, alcohol reduces cancer rates [8].

Arsenic and cadmium are potent human carcinogens that are found in soils, plants, and drinking and natural waters world-wide. However, arsenic is effective in treating leukaemia [9], and low concentrations of cadmium reduce cancer rates to below those in controls [10]. Benzene is a potent human carcinogen but is a natural constituent of roasted coffee [2].

Rodent bioassays that are based on MTD provide misleading predictions of the effects of natural carcinogens on humans at the low concentrations found in food and water. About one-half of all tested synthetic chemicals are also carcinogenic in rodents at MTD [2]. Thus, the identification of such synthetic chemicals as potential carcinogens at the low concentrations normally found in food must be as uncertain as the categorisation of low concentrations of naturally occurring products as harmful. Ames and Gold [11] suggest the rodent test is misleading because high concentrations of many chemicals invariably induce cell division. There is a strong correlation between the induction of cell division at MTD and the subsequent development of cancer, a correlation exemplified by isomers of supposed mutagens in which only the mitogenic isomer induces cancer.

Despite consuming natural carcinogens for thousands of years, humans are not adapted to them. For example, environmental conditions or plant-breeding accidents caused the overproduction of solanine (a fat-soluble nerve toxin) in two lines of potato and of cucurbitacin (now used as an organic insecticide) in organic courgettes. Those that ate the produce experienced intestinal problems and sickness [12,13]. Likewise, the over-production of

psoralen in celery caused extensive skin burns in those who harvested this crop [2].

The concept of hormesis

Hormesis is an unanticipated or paradoxical effect of a toxic chemical(s) or of radiation at low doses. Hundreds of chemicals that have damaging effects at high concentrations paradoxically have beneficial effects on growth, reproduction or longevity, or potentiate immune responses, at low concentrations [14[•],15^{••},16^{••}]. Surveys of both past scientific literature [17] and more recent literature [18] have turned up many hundreds of hormetic dose–response curves. These responses — which are found in plants, animals and bacteria — are commonly described as U- or J-shaped (Figure 1) and are invoked by chemicals that range from toxic metals, insecticides (pesticides) and fungicides, to herbicides and petroleum fractions [16^{••},17,18]. Clearly the natural carcinogens described above are producing hormetic responses.

At low concentrations, environmental dioxins reduce the incidence of human cancers [19]. Contaminated sea-water or diluted factory effluent containing, for example, cadmium and mercury can substantially promote hydroid and algal growth, but only when these elements are present at low concentrations [16^{••},20]. Both selenium deficiency and selenium excess can promote tumour growth, but

selenium is an essential element for human health at the correct concentration [21]. Many other essential minerals and vitamins have similar dose–response characteristics.

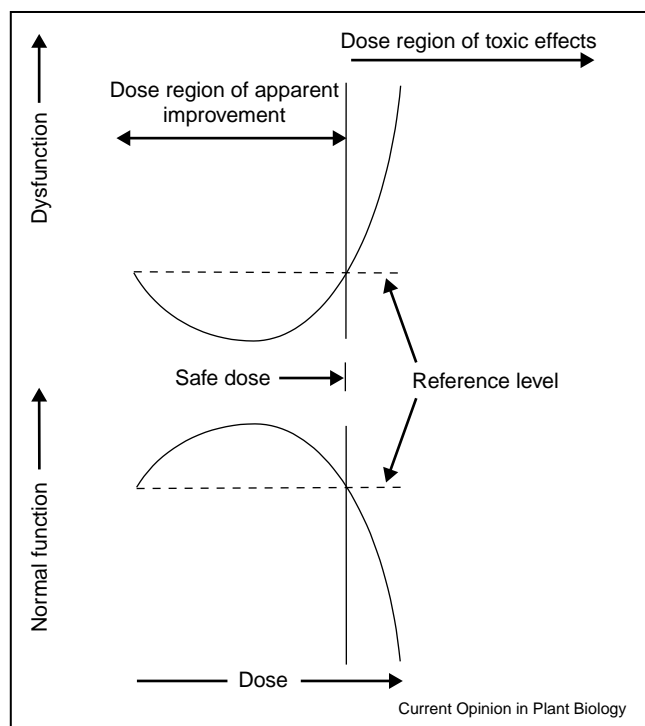
Hormesis is considered to be an over-compensation for a disturbance to homeostasis [22,23]. In analogy, very low UV stress (sunshine) beneficially increases essential vitamin D synthesis; whereas greater, but limited, UV stress stimulates an over-compensation of melanin formation, thereby protecting against subsequent longer UV exposure. UV over-exposure without protection is, however, extremely damaging, causing serious inflammation, accumulations of oxidants and sometimes initiating skin cancer. In the same way, the immune system and DNA-repair systems are potentiated to over-compensate by low levels of chemical stress. In turn, a potentiated immune system contributes to lower cancer rates. Nevertheless, high levels of chemical stress overwhelm homeostasis and induce inflammation, cell division and eventual cancer.

Hormesis brings current US Environmental Protection Agency (EPA) regulatory assessments of synthetic agricultural chemicals [11,24,25] into serious question as low doses can be beneficial to health [15^{••},16^{••}]. Hormesis strikingly contradicts EPA assumptions of ‘no safe dose’ for synthetic chemicals. Not unsurprisingly, early human ancestors evolved a generalised defence mechanism against low levels of ‘toxic’ chemicals to enable their consumption of many different plants containing variable levels of natural carcinogens without subsequent ill-health. Traces of synthetic pesticides (and perhaps mycotoxins) together with the natural pesticides (carcinogens) that are found in fruit and vegetables may potentiate the immune system, helping to protect against cancer. Attempts to clean food of all synthetic chemicals may be counterproductive.

‘Antioxidants’ in fruit and vegetables: protection against the accumulation of mutations

In human adults, superoxide, hydrogen peroxide and hydroxide radicals (produced by some 10^{10} free radicals per cell per day) cause in the order of 10^6 mutational alterations of DNA per cell each day. The activities of these mutagens are countered by ‘antioxidants’, DNA repair, the removal of persistent alterations by apoptosis, differentiation, necrosis and the immune system so that only about one mutation per cell per day persists [25]. Ageing and its associated degenerative diseases — cancer, cataracts and circulation disorders — result in part from oxidative damage to DNA, lipids and proteins. Radiation and serious inflammation also increase oxidant production. By old age, many mutations have accumulated because the repair system is not perfect. The question is not why cancer occurs, but why it occurs so infrequently.

Figure 1



J-shaped dose response curves illustrating an apparent reduction in dysfunction (such as cancer rate) or an improvement in function (such as growth or reproduction) at low doses.

Dietary antioxidant defences are thought to reside in vitamins C and E, quercetin, zeaxanthin and carotenoids such as lycopene. Increased consumption of these antioxidants can reduce DNA oxidation [26]. Some large β -carotene supplementation studies have shown, however, that the incidence of lung cancer rose following increased intake of this antioxidant [27*,28]. Apoptosis may be induced by arsenic and *S*-allylcysteine, a constituent of garlic [29].

Many antioxidants influence other metabolic events. At high doses, vitamin C can be a pro-oxidant [30] but dietary supplementation of this vitamin has been reported to reduce the severity of cardiovascular disease [31*]. The antioxidant capsaicin (from chilli peppers) binds to a receptor in sensitive neurons, thereby elevating their permeability to calcium ions. Prolonged or repeated exposure to capsaicin leads to nerve damage and/or death [32]. The antioxidant glucosinolates (in *Cruciferae*) and their metabolites ([iso]thiocyanates, indoles, epithionitriles and dithiolthiones) exhibit not only anticancer properties but also antibacterial and goitrogenic activities [33,34]. These activities are possibly mediated by cytochrome p450 enzymes, which activate nitrosamines that alkylate carcinogens.

The more classical antioxidants — anthocyanins, procyanidins (tannins), flavonoids, hydroxy benzoic acid (HBA) derivatives and so on — inhibit the oxidation of low-density lipoproteins, transcriptionally reduce the synthesis of the vaso-constrictive peptide endothelin-1, and reduce platelet aggregation (by inhibiting cyclooxygenases and lipoxygenases) with a subsequent retardation of atherosclerosis [35,36]. Many classical antioxidants also exhibit the chelation of metal ions, thereby impairing mineral intake. Some flavonoids are reported to exhibit anti-ulcer/gastroprotective effects by inhibition of acid secretion and/or elevation of prostaglandin, leading to elevated duodenal bicarbonate secretion, and also have antibacterial action against *Helicobacter pylori* [36,37]. Whereas some flavonoids protect against some cancers [38], others (e.g. genistein) can negate the inhibitory effects of chemotherapy on cancerous growth [39]. Small phenolics, such as gallic, gentisic and salicylic acids, inhibit cancer in cell-line studies. The acetylation of salicylic acid to make aspirin greatly increases its uptake, changing its dose–response characteristics. The actual level of these compounds in the plasma may not, however, equate to their consumption level as the gut microflora are known to degrade other phenolics, such as cinnamates and flavonoids, to hydroxy-benzoic acids.

Other recently discovered natural products in foodstuffs, such as stanol esters, effectively lower serum cholesterol. These compounds do not accumulate in plasma themselves, but feruyolated stanols and sterols in corn oil may provide dietary benefits both by this effect on chole-

sterol levels and through antioxidant protection [40]. Soluble extracts of bitter melon or ginger rhizome inhibit mammary carcinogenesis [41] but the active chemicals remain unknown. The extent to which a diet that is high in any of these chemicals reduces oxidant damage to DNA has yet to be determined. The alternative, potentiation of DNA repair needs further investigation, particularly as cell oxidant concentrations may be homeostatically regulated.

Carcinogenic mycotoxins

The fungal mycotoxins aflatoxin, ochratoxin, patulin and fumonisin, and the tricothrones, are produced by species of *Penicillium*, *Fusarium* and *Aspergillus* growing on a wide variety of foods. All these mycotoxins can act as carcinogens in either rodents or humans; in addition, they weaken the immune system, exposing damaged individuals to other diseases [42,43,44*]. Economic loss from the contamination of food by these mycotoxins is substantial: estimated to exceed ten billion US dollars in the US alone [45]. Aflatoxin is the most potent human carcinogen, inducing aflatoxicosis and liver cancer within several months; its effects on poultry can be detected in the parts per billion range. The consumption of all mycotoxins by humans is currently considered to be inevitable. It is thought that present ‘safe’ levels might induce cancer in a minority of individuals even in western countries [45]. If the dose response is hormetic, and this is currently unknown, then safe levels might actually improve health. A need for more research is clearly indicated.

Mycotoxic fungi infect both grains and fruit; their growth is encouraged by poor storage conditions. Pre-harvest pest or mechanical damage to cereal grains, cobs and fruits enables fungal spores to enter the plant tissues, and these spores germinate when the crop is moist. Post-harvest fungicide treatments are used to control fungal infection but penetration may limit the effectiveness of these treatments. Furthermore, post-harvest pesticides are the major source of synthetic pesticide traces in food [46]. Early results indicate that expression of the *Bacillus thuringiensis* toxin (Bt) protein in corn, reduced both pre-harvest pest damage and contamination by fumonisin and aflatoxin by 5–30 fold [44*]. Harvesting in wet conditions and storage of seed at warm temperatures encourage fungal growth, and rapid drying is necessary to discourage contamination [47]. Small farms often lack the necessary expensive drying equipment, and this might contribute to the frequent reports of higher fumonisin contamination of organic wheat (e.g. [48–50]), as organic associations frequently emphasise the virtues of small farms. In dry years, fumonisin levels seem unrelated to the mode of farming [51]. Human vaccination against aflatoxin is being used to combat this problem [43], and the construction of crops that are transformed with fungal genes that degrade fumonisin are an alternative solution [52].

Contamination of peanuts by aflatoxin is encouraged by severe drought followed by heat stress during fruit development, which compromise plant defence responses and increase the risk of infection [53]. Placing a battery of disease resistance genes under transcriptional control by abscisic acid or heat shock promoters might help reduce aflatoxin contamination. Fungal contamination of peanuts may also result in more serious aflatoxin contamination during the preparation of peanut products and in other nuts, such as walnuts, hazelnuts and so on, that are poorly dried during harvest.

Patulin is synthesised in contaminated apples that are infected by *Penicillium* species [54,55]. Even fruit with minor spots of infection or no apparent blemishes can be contaminated throughout, suggesting that *Penicillium* species may be endemic in apple trees because all are vegetatively propagated. Patulin concentrations have been reported to be higher in organic apple juice. The reasons for this are not clear [49] but the lack of use of effective fungicides might be responsible. At present, patulin contents generally appear to be within safety guidelines for consumption [56].

Deficiencies in diet

About 80% of US and 75% of UK citizens eat insufficient fruit and vegetables to provide minimal protection against cancer [2]. Sufficient, and sometimes excessive, calories are consumed but the diet is imbalanced as regards the necessary components from fruit and vegetables. Vitamin A deficiency in early childhood potentiates visual disorders and sensitivity to childhood diseases that can lead to death. Genetically modified vitamin-A-enhanced rice is currently being researched in field trials because rice gruel, a commonly used substitute for breast milk, is severely deficient in the vitamin A necessary to synthesise retinal, the dominant visual pigment. Many necessary nutrients exhibit a hormetic dose response, however, and vitamin A in excess can cause bone disorders and cancer. A common deficiency in folic acid reduces mitotic rates (particularly in bone marrow, leading to anaemia) and enhances the replacement of thymine by uracil during DNA replication, potentially causing chromosome breakage and eventual cancer [21]. Folate deficiency accelerates homocysteine accumulation, thereby damaging endothelial cells in culture, and is a risk factor for arterial endothelial dysfunction.

Conclusions

The inhibition of cancer, cardiovascular disease and other degenerative diseases is the biggest goal facing nutritional plant breeding [57]. The potential for genetic modification of dietary chemicals is substantial, but this short review indicates the difficulties of assuming that increasing the dietary level of any compound will necessarily improve health. Many plant secondary products and dietary contaminants seem to have paradoxical (hormetic)

effects on these diseases that depend on their concentration and thus level of consumption. For the individual, the conclusion must be that everyone should eat more fruits and vegetables but must live with potential risks, particularly as there is likely to be variation amongst the human population in sensitivity to many of the natural and supposedly hazardous chemicals that these foods contain. More dramatically, the consumption of conventionally grown fruits and vegetables that containing traces of pesticides that have hormetic properties should involve lower health risks than the consumption of organic fruits and vegetables in which these products have purportedly been eliminated. This contradicts common assumptions about toxicity, such as those made by Baker *et al.* [46]. Organic food that is contaminated with varying amounts of the organic pesticides copper or rotenone provides further problems for the assumption that organic is necessarily better [58].

A change in EPA regulations with regard to chemicals that have pronounced hormetic properties must be a priority. Re-educating the public, who have been told for so long that a toxin is toxic, no matter the concentration, will not be easy but should be the goal of any scientist interested in the health of the community. A balanced conventional diet is currently the most promising route to healthy eating, particularly as the 200 investigations summarised by Block *et al.* [1] to indicate that a diet that is high in fruit and vegetables cuts cancer risks substantially used conventional fruits and vegetables rather than organic ones. For the food scientist, there is a clear need to generate detailed information on the metabolomics of both fruits and vegetables that have been grown and stored under many different conditions, and to produce hormetic dose-response data for many of the compounds contained in these foods. The manipulation of dietary constituents by genetic modification can then proceed with confidence.

References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
 - of outstanding interest
1. Block G, Patterson B, Subar A: **Fruit vegetables and cancer prevention. A review of the epidemiological evidence.** *Nutr Cancer* 1992, **18**:1-29.
 2. Ames BN, Gold LS: **Paracelsus to parascience: the environmental cancer distraction.** *Mutat Res* 2000, **447**:3-13.
 3. Eberhardt MV, Lee CY, Liu RH: **Antioxidant activity of fresh apples.** *Nature* 2000, **405**:903-904.
 4. Ranelletti FO, Maggiano N, Serra FG, Ricci R, Larocca LM, Lanza P, Scambia G, Fattorossi A, Capelli A, Piantelli M: **Quercetin inhibits p21-ras expression in human colon cancer cell lines and in primary colorectal tumors.** *Int J Cancer* 2000, **85**:438-445.
 5. Stark MJ, Burke YD, McKinzie JH, Ayoubi AS, Crowell PL: **Chemotherapy of pancreatic cancer with the mono-terpene family perillyl alcohol.** *Cancer Lett* 1995, **96**:15-21.
 6. Kopp-Schneider A, Lutz WK: **J shaped dose response relationship for tumour induction by caffeic acid in the rat fore**

- stomach, modeled by non monotonic dose response for DNA damage and cell proliferation. *Human Ecol Risk Assess* 2001, 7:921-931.
7. Conaway CC, Yang YM, Chung FL: **Isothiocyanates as cancer chemo-protective agents: their biological activities and metabolism in rodents and humans.** *Curr Drug Metab* 2002, 3:233-255.
 8. Rehm J, Greenfield TK, Rogers JD: **Average volume of alcohol consumption, patterns of drinking and all-cause mortality: results from the US national alcohol survey.** *Am J Epidemiol* 2001, 153:64-71.
 9. Munshi NC, Tricot G, Desikan R, Badros A, Zangari M, Toor A, Morris C, Anaissie E, Barlogie B: **Cadmium trioxide for the treatment of multiple myeloma.** *Leukemia* 2002, 16:1835-1837.
 10. Waalkes MP, Rehm S, Riggs CW, Bare CM, Devor DE, Poirier LA, Wenk ML, Henneman JR, Balasak MS: **Cadmium carcinogenesis in male Wistar CrI(WI)BR/ rats: dose response analysis of tumour induction in the prostate and testes at the injection site.** *Cancer Res* 1998, 48:4656-4663.
 11. Ames BN, Gold LS: **Pollution pesticides and cancer misconceptions.** In *Fearing Food: Risk, Health and Environment*. Edited by Morris J, Bate R. Oxford: Butterworth-Hienemann; 1999:19-38.
 12. Van Gelder WMJ, Vinke JH, Scheffer JJC: **Steroidal glycoalkaloids in tubers and leaves of solanum species used in potato breeding.** *Euphytica* 1988, 88:147-158.
 13. Anon: **Food safety advice. Toxins in courgettes.** *Food Safety Quarterly Report* 2001. Auckland: Auckland District Health Board; 2001.
 14. Calabrese EJ, Baldwin LA: **U shaped dose responses in biology, • toxicity and health.** *Annu Rev Public Health* 2001, 22:15-33. A valuable introduction to the history of hormesis and a simple introduction to the whole subject of hormesis.
 15. Calabrese EJ, Baldwin LA: **U shaped dose responses and their • centrality in toxicology.** *Trends Pharmacol Sci* 2001, 22:285-291. One of two crucial articles [15**,16**] for anyone wishing to avail themselves of the recent and previous literature on hormesis. This review was written by two authors who have been instrumental in recovering the concept of hormesis from the doldrums into which it had entered. The article describes some of the many hundreds of paradoxical responses of both plants and animals to toxic chemicals.
 16. Calabrese EJ, Baldwin LA: **Applications of hormesis in • toxicology, risk assessment and chemotherapeutics.** *Trends Pharmacol Sci* 2002, 23:331-337. The second of two articles [15**,16**] that point to the importance of hormesis in changing the perspective from which all toxic chemicals are assessed in various fields of study. This article also contains dose-response data for the effluent from two kinds of industrial factory, indicating how such pollution can be used productively to increase algal growth.
 17. Calabrese EJ, Baldwin LA: **Hormesis as a biological hypothesis.** *Environ Health Perspect* 1998, 106(Suppl 1):357-362.
 18. Calabrese EJ, Baldwin LA: **Hormesis — a generalisable and unifying hypothesis.** *Crit Rev Tox* 2001, 31:353-424.
 19. Andersen ME, Conolly RB: **Mechanistic modeling of rodent liver tumor promotion at low levels of exposure: an example related to dose response relationships for 2,3,7,8 tetrachlorodibenzo-p-dioxin.** *Human Exptl Toxicol* 1998, 17:683-690.
 20. Stebbings ARD: **A theory for growth hormesis.** *Mutat Res* 1998, 403:239-258.
 21. Ames BN: **DNA damage from micronutrient deficiencies is likely to be a major cause of cancer.** *Mutat Res* 2001, 475:7-20.
 22. Calabrese EJ: **Overcompensation stimulation: a mechanism for hormetic effects.** *Crit Rev Toxicol* 2001, 31:425-470.
 23. Deng C, Grahma R, Shukla R: **Detecting and estimating hormesis using a model based approach.** *Human Ecol Risk Assess* 2001, 7:849-866.
 24. Kodell R: **U shaped dose response relationships for mutation and cancer.** *Human Ecol Risk Assess* 2001, 7:909-919.
 25. Polycove M, Feinendegen LE: **Molecular biology, epidemiology and the demise of the linear no-threshold (LNT) hypothesis.** *Compt Rend Acad Sci Serie III-Life Sci* 1999, 322:197-204.
 26. Dusinka M, Vallova B, Ursinyova M, Hladikova V, Smolkova B, Wsolova L, Raslova K, Collins AR: **DNA damage and antioxidants; fluctuations through the year in a central European population group.** *Food Chem Technol* 2002, 40:1119-1123.
 27. Holick CN, Michaud DS, Stolzenberg-Solomon R, Mayne ST, Pietinen P, Taylor PR, Virtamo J, Albanes D: **Dietary carotenoids, serum beta-carotene, and retinol and risk of lung cancer in the alpha-tocopherol, beta-carotene cohort study.** *Am J Epidemiol* 2002, 156:536-547. This paper describes the results of the beta-Carotene and Retinol Efficacy Trial (CARET), which studied the effect of carotenoid supplementation on the incidence of lung cancer in more than 18 000 people. Supplementation actually caused slight increases in lung cancer rates.
 28. Omenn GS, Goodman GE, Thornquist MD, Balmes J, Cullen MR, Glass A, Keogh JP, Meyskens FL, Valanis B, Williams JH et al.: **Effects of a combination of beta carotene and vitamin A on lung cancer and cardiovascular disease.** *N Engl J Med* 1996, 334:1150-1155.
 29. Balasenthil S, Ra KS, Nagini S: **Apoptosis induction by S-allylcysteine a garlic constituent during 7,12 dimethylbenz[a]anthracene-induced hamster buccal pouch carcinogenesis.** *Cell Biochem Funct* 2002, 20:263-268.
 30. Podmore ID, Griffiths HR, Herbert KE, Mistry N, Mistry P, Lunec J: **Vitamin C exhibits pro-oxidant properties.** *Nature* 1998, 392:559-560.
 31. Josphipura KJ, Hu FB, Manson JE, Stampfer MJ, Rimm EB, Speizer • FE, Colditz G, Ascherio A, Rosner B, Spiegelman D, Willett WC: **The effect of fruit and vegetable intake on risk for coronary heart disease.** *Ann Intern Med* 2001, 134:1106-1114. A significant and in-depth study of the relationship between diet and disease in more than 80 000 women and more than 40 000 men over 14 and 8 years, respectively. Supplementation reduced cardiovascular disorders.
 32. Buck SH, Burks TF: **The neuropharmacology of capsaicin — review of some recent observations.** *Pharmacol Rev* 1986, 38:179-226.
 33. Van Poppel G, Verhoeven DTH, Verhagen H, Goldbohm RA: **Brassica vegetables and cancer prevention: epidemiology and mechanisms.** *Adv Exp Med Biol* 1999, 472:159-168.
 34. Mithen RF, Dekker M, Verkerk R, Rabot S, Johnson IT: **The nutritional significance, biosynthesis and bioavailability of glucosinolates in human foods.** *J Sci Food Agric* 2000, 80:967-984.
 35. Corder R, Douthwaite JA, Lees DM, Khan NQ, Santos ACV, Wood EG, Carrier MJ: **Endothelin-1 synthesis reduced by red wine.** *Nature* 2001, 414:863.
 36. Hollman PCH, Katan MB: **Dietary flavonoids: intake, health effects and bioavailability.** *Food Chem Technol* 1999, 37:937-942.
 37. Martin ML, Alarcón de la Lastra C, Motilva V, La Casa C: **Antiulcer and gastroprotective activity of flavonic compounds: mechanisms involved.** In *Studies in Natural Product Chemistry*. Edited by Rahman AU. Amsterdam: Elsevier Science; 2000:419-456.
 38. Knekt P, Järvinen R, Seppänen R, Heliövaara M, Teppo L, Pukkala E, Aromaa A: **Dietary flavonoids and the risk of lung cancer and other malignant neoplasms.** *Am J Epidemiol* 1997, 146:223-230.
 39. Ju YH, Doerge DR, Allred KF, Allred CD, Helferich WG: **Dietary genistein negates the inhibitory effect of Tamoxifen on growth of estrogen-dependent human breast cancer (MCF-7) cells implanted in athymic mice.** *Cancer Res* 2002, 62:2474-2477.
 40. Ramjiganesh T, Roy S, Nicolosi RJ, Young TL, McIntyre JC, Fernandez ML: **Corn husk oil lowers plasma LDL cholesterol concentrations by decreasing cholesterol absorption and altering hepatic cholesterol metabolism in guinea pigs.** *J Nutr Biochem* 2000, 11:358-366.
 41. Nagasawa H, Watanabe K, Inatomi H: **Effects of bitter melon or ginger rhizome on spontaneous mammary tumorigenesis in SHN mice.** *Am J Chin Med* 2002, 30:195-205.

42. Bondy GS, Pestka JJ: **Immunomodulation by fungal toxins.** *J Toxicol Environ Health B Critical Rev* 2000, **3**:109-143.
43. Henry SH, Bosch FX, Troxell TC, Bolger PM: **Reducing liver cancer — aflatoxin.** *Science* 1999, **286**:2453-2454.
44. Minorsky PV: **Fumonisin mycotoxins.** *Plant Physiol* 2002, **129**:929-930.
 An excellent short review on fumonisin mycotoxins that includes a description of how the concentrations of fumonisin are reduced in plants that are transformed to contain the *Bacillus thuringiensis* toxin (Bt) gene.
45. Miller JD: **Global significance of mycotoxins.** In *Mycotoxins and Phycotoxins — Development, Chemistry, Toxicology and Food Safety*. Edited by Miraglia M, van Egmond H, Brera C, Gilbert J. Colorado: Alken Inc.; 2000:3-17.
46. Baker BP, Benbrook CM, Groth E, Benbrook KL: **Pesticide residues in conventional, integrated pest management grown and organic foods: insights from three US data sets.** *Food Addit Contam* 2002, **19**:427-446.
47. Birzele B, Prange A, Kramer J: **Deoxynivalenol and ochratoxin A in German wheat and changes of level in relation to storage parameters.** *Food Addit Contam* 2000, **17**:1027-1035.
48. Czerwiecki L, Czajkowska D, Witkowska-Gwiazdowska A: **On ochratoxin A and fungal flora in Polish cereals from conventional and ecological farms.** *Food Addit Contam* 2002, **19**:470-477.
49. Malmauret L, Parent-Massin D, Hardy JL, Verger P: **Contaminants in organic and conventional foodstuffs in France.** *Food Addit Contam* 2002, **19**:524-532.
50. Kirchmann H, Thorvaldsson G: **Challenging targets for future agriculture.** *European J Agron* 2000, **12**:145-161.
51. Schollenberger M, Jara HT, Suchy S, Drochner W, Muller HM: **Fusarium toxins in wheat flour collected in an area of South West Germany.** *Int J Food Microbiol* 2002, **72**:85-89.
52. Duvick J: **Prospects for reducing fumonisin contamination of maize through genetic modification.** *Environ Health Perspect* 2001, **109**(Suppl 2):337-342.
53. Cole RJ, Dörner JW, Blankenship PD: **Management strategies for prevention and control of mycotoxins.** In *Mycotoxins and Phycotoxins — Development, Chemistry, Toxicology and Food Safety*. Edited by Miraglia M, van Egmond H, Brera C, Gilbert J. Colorado: Alken Inc.; 1998:189-203.
54. Martins ML, Gimeno A, Martins HM, Bernardo F: **Co-occurrence of patulin and citrinine in Portuguese apples with rotten spots.** *Food Additives Contam* 2002, **19**:568-574.
55. Kadakal C, Nas S: **Effect of apple decay proportion on the patulin, fumaric acid, HMF and other apple juice properties.** *J Food Safety* 2002, **22**:17-25.
56. Lai CL, Fuh YM, Shih DY: **Detection of mycotoxin patulin in apple juice.** *J Food Drug Anal* 2000, **8**:85-90.
57. Parr AJ, Bolwell GP: **Phenols in the plant and in man. The potential for possible nutritional enhancement of the diet by modifying the phenols content or profile.** *J Sci Food Agric* 2000, **80**:985-1012.
58. Trewavas AJ: **Urban myths of organic farming.** *Nature* 2001, **410**:409-410.